

FYL- U5

FYL Rapid Test Single Panel (Urine)

For forensic use only.

INTENDED USE

The FYL Rapid Test Single Panel (Urine) is a rapid visual immunoassay for the qualitative, presumptive detection of FYL in human urine specimens at the cut-off concentrations listed below:

 Parameter
 Calibrator
 Cut-off (ng/mL)

 FYL
 Fentanyl
 200

INTRODUCTION

Fentanyl is a synthetic opioid related to the phenylpiperidines. Fentanyl is approximately 100 times more potent than morphine. This agent is highly lipid soluble and rapidly cross the blood-brain barrier. This is reflected in the half-life for equilibration between the plasma and cerebrospinal fluid of approximately 5 minutes for fentanyl. The levels in plasma and cerebrospinal fluid decline rapidly owing to redistribution of fentanyl from highly perfused tissue groups to other tissues, such as muscle and fat. As saturation of less well-perfused tissue occurs, the duration of effect of fentanyl and sufentanii approaches the length of their elimination half-lives of between 3 and 4 hours. Fentanyl undergoes hepatic metabolism and renal excretion. Therefore, with the use of higher doses or prolonged infusions, fentanyl becomes longer acting.

PRINCIPLE

The FYL Rapid Test Single Panel (Urine) detects FYL through visual interpretation of color development on the Single Panel. Drug conjugates are immobilized on the test region of the membrane. During testing, the specimen reacts with antibodies conjugated to colored particles and precoated on the sample pad. The mixture then migrates through the membrane by capillary action, and interacts with reagents on the membrane. If there are insufficient drug molecules in the specimen, the antibody-colored particle conjugate will bind to the drug conjugates, forming a colored band at the test region of the membrane. Therefore, a colored band appears in the test region when the urine is negative for the drug. If drug molecules are present in the urine above the cut-off concentration of the test, they compete with the immobilized drug conjugate on the test region for limited antibody binding sites. This will prevent attachment of the antibody-colored particle conjugate to the test region. Therefore, the absence of a colored band at the test region indicates a positive result. The appearance of a colored band at the control region serves as a procedural control, indicating that the proper volume of specimen has been added and membrane wicking has occurred.

REAGENTS

Each test consists of a reagent Single Panel mounted in a plastic housing. The amount of each antigen and/or antibody coated on the Single Panel is less than 0.001 mg for antigen conjugates and goat anti-rabbit IgG antibodies, and less than 0.0015 mg for antibody components.

The control zone of each test contains goat anti-rabbit IgG antibody. The test zone of each test contains drug-bovine protein antigen conjugate, and the conjugate pad of each test contains monoclonal anti-drug antibody and rabbit antibody-colored particle complex.

MATERIALS

Materials Provided

Test Panel
 Package insert

Materials Required but Not provided

Positive and negative controls

Time

Centrifuge

PRECAUTIONS

- For forensic use only.
- Do not use after the expiration date indicated on the package. Do not use the test if the foil pouch or canister is damaged. Do not reuse tests.
- This kit contains products of animal origin. Certified knowledge of the origin and/or sanitary state
 of the animals does not completely guarantee the absence of transmissible pathogenic agents. It is
 therefore, recommended that these products be treated as potentially infectious, and handled by
 observing usual safety precautions (e.g., do not ingest or inhale).
- Avoid cross-contamination of specimens by using a new specimen collection container for each specimen obtained.
- Read the entire procedure carefully prior to testing.
- Do not eat, drink or smoke in the area where specimens and kits are handled. Handle all specimens
 as if they contain infectious agents. Observe established precautions against microbiological hazards
 throughout the procedure and follow standard procedures for the proper disposal of specimens.
 Wear protective clothing such as laboratory coats, disposable gloves and eye protection when
 specimens are assayed.
- Humidity and temperature can adversely affect results.

· Used testing materials should be discarded in accordance with local regulations.

STORAGE AND STABILITY

- The kit should be stored at 2-30°C until the expiry date printed on the sealed pouch or canister.
- The test must remain in the sealed pouch or closed canister until use.
- Do not freeze.
- Kits should be kept out of direct sunlight.
- Care should be taken to protect the components of the kit from contamination. Do not use if there is
 evidence of microbial contamination or precipitation. Biological contamination of dispensing
 equipment, containers or reagents can lead to false results.

SPECIMEN COLLECTION AND STORAGE

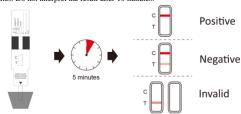
- . The FYL Rapid Test Single Panel (Urine) is intended for use with human urine specimens only.
- Urine collected at any time of the day may be used.
- · Urine specimens must be collected in clean, dry containers.
- Turbid specimens should be centrifuged, filtered, or allowed to settle and only the clear supernatant should be used for testing.
- Perform testing immediately after specimen collection. Do not leave specimens at room temperature for prolonged periods. Urine specimens may be stored at 2-8°C for up to 2 days. For long term storage, specimens should be kept below -20°C.
- Bring specimens to room temperature prior to testing. Frozen specimens must be completely thawed
 and mixed well prior to testing. Avoid repeated freezing and thawing of specimens.

If specimens are to be shipped, pack them in compliance with all applicable regulations for transportation of etiological agents.

PROCEDURE

Bring tests, specimens, and/or controls to room temperature (15-30 $^{\circ}$ C) before use it if the specimens are refrigerated.

- Remove the test from its sealed pouch and use it as soon as possible. To obtain a best result, the assay should be performed within one hour.
- Hold the Single Panel at the handle with the product name imprints. Do not touch the membrane
 part of the Single Panel to avoid contamination.
- Dip the test Single Panel vertically in the urine specimen for at least 10-15 seconds. Do not pass
 the maximum line (MAX) on the test Single Panel when immersing the Single Panel. As the test
 begins to work, you will see color move across the membrane.
- 4. Take the Single Panel out of the specimen afterwards and place it on a non-absorbent flat surface. Start the timer and wait for the colored line(s) to appear. The result should be read at 5 minutes. Do not interpret the result after 10 minutes.



INTERPRETATION OF RESULTS

POSITIVE: Only one colored band appears, in the control region (C). No

apparent colored band appears in the test region (T).

NEGATIVE: Two colored bands appear on the membrane. One band appears in the control region (C) and another band appears in the test region (T)

INVALID: Control band fails to appear. Results from any test which has not produced a control band at the specified read time must be discarded. Please review the procedure and repeat with a new test. If the problem persists, discontinue using the kit immediately and contact your local distributor.

NOTE:

- The intensity of color in the test region (T) may vary depending on the concentration of analytes present
 in the specimen. Therefore, any shade of color in the test region should be considered negative. Note that
 this is a qualitative test only, and cannot determine the concentration of analytes in the specimen.
- Insufficient specimen volume, incorrect operating procedure or expired tests are the most likely reasons for control band failure

OUALITY CONTROL

· Internal procedural controls are included in the test. A colored band appearing in the control region

- (C) is considered an internal positive procedural control, confirming sufficient specimen volume and correct procedural technique.
- External controls are not supplied with this kit. It is recommended that positive and negative
 controls be tested as a good laboratory practice to confirm the test procedure and to verify proper
 test performance.

LIMITATIONS OF THE TEST

- The FYL Rapid Test Single Panel (Urine) is for forensic use only, and should be only used for the qualitative detection of FYL.
- 2. This assay provides a preliminary analytical test result only. A more specific alternative chemical method must be used in order to obtain a confirmed analytical result. Gas chromatography/mass spectrometry (GC/MS) has been established as the preferred confirmatory method by the National Institute on Drug Abuse (NIDA). Clinical consideration and professional judgment should be applied to any test result, particularly when preliminary positive results are indicated.
- There is a possibility that technical or procedural errors as well as other substances and factors may interfere with the test and cause false results.
- Adulterants, such as bleach and/or alum, in urine specimens may produce erroneous results regardless
 of the analytical method used. Therefore, please preclude the possibility of urine adulteration prior to
 testine.
- 5. A positive result indicates the presence of a FYL only, and does not indicate or measure intoxication.
- A negative result does not at any time rule out the presence of FYL in urine, as they may be present below the minimum detection level of the test.
- 7. This test does not distinguish between FYL and certain medications.

PERFORMANCE CHARACTERISTICS

The accuracy of the FYL Rapid Test Single Panel (Urine) was compared and checked against commercially available tests with a threshold value at the same cut-off levels. Urine samples taken from volunteers claiming to be non-users were examined under both tests. The results were >95.7% in agreement.

B. Reproducibility

A. Accuracy

The reproducibility of the FYL Rapid Test Single Panel (Urine) was verified by blind tests performed at four different locations. Samples with FYL concentrations at 50% of the cut-off were all determined to be negative, while samples with FYL concentrations at 200% of the cut-off were all determined to be positive.

C. Precision

Test precision was determined by blind tests with control solutions. Controls with FYL concentrations at 50% of the cut-off yielded negative results, and controls with FYL concentrations at 150% of the cut-off yielded positive results.

D. Specificity

The following tables list the concentrations of compounds (ng/mL) above which the FYL Rapid Test Single Panel (Urine) identified positive results at 5 minutes.

FYL 200 related compounds

Fentanyl and Fentanyl metabolites	20
Fentanyl	200
Norfentanyl	>10,000

The following compounds yielded negative results up to a concentration of 100 µg/mL

The following compounds yielded negative results up to a concentration of 100 µg/m2.			intration of 100 µg/mil.
	(-)-Ephedrine	Chlorpheniramine	Oxalic Acid
	(+)-Naproxen	Creatine	Penicillin-G
	(+/-)-Ephedrine	Dextromethorphan	Pheniramine
	4-Dimethyllaminoantiyrine	Dextrorphan tartrate	Phenothiazine
	Acetaminophen	Dopamine	Procaine
	Acetone	Erythromycin	Protonix
	Albumin	Ethanol	Pseudoephedrine
	Amitriptyline	Furosemide	Quinidine
	Ampicillin	Glucose	Ranitidine
	Aspartame	Guaiacol Glyceryl Ether	Sertraline
	Aspirin	Hemoglobin	Tyramine
	Benzocaine	Ibuprofen	Vitamin C (Ascorbic Acid)
	Bilirubin	Imipramine	Trimeprazine
	b-Phenylethyl-amine	Isoproterenol	Venlafaxine
	Caffeine	Lidocaine	Methadone
	Chloroquine		

LITERATURE REFERENCES

- Baselt RC. Disposition of Toxic Drugs and Chemicals in Man. 2nd ed. Davis: Biomedical Publications; 1982.
- Hawks RL, Chiang CN, eds. Urine Testing for Drugs of Abuse. Rockville: Department of Health and Human Services, National Institute on Drug Abuse; 1986.
- Substance Abuse and Mental Health Services Administration. Mandatory Guidelines for Federal Workplace Drug Testing Programs. 53 Federal Register; 1988.
- McBay AJ. Drug-analysis technology--pitfalls and problems of drug testing. Clin Chem. 1987 Oct; 33 (11 Suppl): 33B-40B.
- 5. Gilman AG, Goodman LS, Gilman A, eds. Goodman and Gilman's The Pharmacological Basis of

Therapeutics. 6th ed. New York: Macmillan; 1980

GLOSSARY OF SYMBOLS

٥	Catalog number	8	Temperature limitation
ι	Consult instructions for use	Λ	Batch code
I	In vitro diagnostic medical device	3	Use by
и	Manufacturer	σ	Do not reuse

Manufactured for: **American Screening LLC** 9742 St. Vincent Ave., Suite 100 Shreveport, Louisiana 71106

Customer Service Phone: 866-526-2873



American Screening LLC 9742 St. Vincent AVE Suite 100 Shreveport, LA 71105 1-318-798-3306

IFU 21-9030, Rev. A Number:1110029030

REV1.0/Effective date:2019-08-07